LLT.A. Accession line: Inoculum TVn 32 TVu 37 TVu 76 TVu 57 TVu 317 TVu 400 TVu 2366 TVn 2398 Colletotrichum lindemuthianum (1) (1) (1)(1)isolate I 47 (ex I.I.T.A.) Tobacco necrosis virus **(1**) (2)\*(1) (2)**(1)** (1) (1)

Table 1. Occurrence of 2'-O-methylphaseollidinisoflavan (1) and phaseollidin (2) in infected cowpea stems

287 (3·80) [3]. In view of the reported occurrence of phaseollidin in cowpea line TVu 76 following inoculation with tobacco necrosis virus [1], seven additional lines were examined qualitatively for the presence of (1) and (2), with the results shown in Table 1. Lines TVu 32, 37 and 76 are susceptible [7] to *C. lindemuthianum* and therefore required virus inoculation to give an adequate yield of antifungal compounds; the other lines investigated were classified as resistant. (1) and (2) were identified by TLC (SiO<sub>2</sub>, CHCl<sub>3</sub>:C<sub>2</sub>H<sub>5</sub>OH 97:3 and C<sub>6</sub>H<sub>6</sub>:MeCOOC<sub>2</sub>H<sub>5</sub> 3:1) and by their UV spectra. It is appreciated that mixtures of (1) and (2) might escape identification by these methods if the minor component is present only in trace quantities.

Compound (1) totally inhibited conidiospore germination of Nigerian isolates I 47 and I 57 (ex I.I.T.A.) of *C. lindemuthianum* at 10 and 15 ppm re-

spectively; the values for compound (2) were 20 and 25 ppm.

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## TYRAMINE FROM MAGNOLIA SPECIES

## H. MATSUTANI and T. SHIBA

Fukui Technical College, Sabae, Fukui, 916, Japan and Department of Chemistry, Faculty of Science, Osaka University, Toyonaka, Osaka, 560, Japan

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**Key Word Index**—Magnolia denudata; M. liliiflora; M. obovata; M. kobus; M. grandiflora; Magnoliaceae; amine; tyramine.

In Chinese medicine, Shin-i, prepared from dried young buds of *Magnolia* plants, is used as a sedative or an analgesic. In Japan, Shin-i taken internally for treatment of headaches or colds

are also young buds of *M. kobus* or *M. salicifolia*. In addition, *M. obovata* is utilized in Japan for abdominal distention or pains, and as a diuretic, and *M. grandiflora* for headache or giddiness.

<sup>\*</sup> See Ref. 1.

Table 1. Tyramine content of Magnolia species

Species	Collection time	Plant part	Tyramine content (mg/100 g)
M. denudata	mid September	Leaves	8.0*
M. liliiflora	mid September	Leaves	2.5*
M. liliiflora	late May	Buds	7.8†
M. obovata	late May	Leaves	2.8*

<sup>\*</sup> Method A: content per 100 g of plant parts was calculated based on the weight of crystalline tyramine monohydrochloride isolated.

Investigation of leaves, buds and flowers of 5 Magnolia species revealed the presence of tyramine as a common component besides several of the normal protein amino acids. This unusual occurrence of tyramine, known as an important adrenergic drug [1, 2], may be responsible for the pharmacological action of the oriental folk medicines mentioned above. The tyramine content of the plants examined are shown in Table 1. In addition, qualitative tests on TLC of aqueous extracts from flowers and leaves of M. kobus and leaves of M. grandiflora also showed the presence of tyramine in appreciable amounts.

It was shown that tyramine was not formed by enzymatic decarboxylation of tyrosine during the isolation procedure, since the amounts of the amine did not vary after either immediate boiling the fresh leaves of M. denudata or keeping the homogenate at room temperature for several hours.

The identity of the isolated sample with authentic tyramine monohydrochloride was confirmed by elemental analysis, amino acid analysis (15 cm column, 0·24 N borate buffer of pH 11·3; retention time; 29 min), IR, UV and NMR spectra.

#### EXPERIMENTAL

Isolation of tyramine. The fr. leaves (500 g) of M. obovata collected in the end of May were macerated and extracted  $2\times$  with boiling  $H_2O$ . The combined filtrate (3 l.) was chromatographed on an Amberlite CG-50 (NH<sub>4</sub><sup>+</sup> form) column  $3\times$  50 cm. The cluate with  $2 \text{ N-NH}_4OH$  (0-6 l.) was evaporated in vacuo. Repetition of the column chromatography with gradient elution of NH<sub>3</sub> gave pure tyramine which was crystallized from EtOH as mono HCl-ide (14 mg) as leaflets, mp  $245-255^\circ$  (dec.). The same method was used for the other 2 species. IR,  $\nu$  3180, 1620, 1600 cm<sup>-1</sup>; UV,  $\lambda_{\text{moH-H}_2O}^{\text{HOH-H}_2O}$  282, 276, 223, 198 nm ( $\epsilon$  1420, 1670, 7990, 2770); NMR (D<sub>2</sub>O) 100 MHz,  $\delta$ 7·2 (4H, q, aromatic proton), 3·2 (4H,  $\Delta$ 2B<sub>2</sub> type). Anal. found: C, 54·90, H, 6·96; N, 7-95; Cl, 20·26. Calcd for  $C_8H_{12}$ ONCl: C, 55·34; H, 6·97; N, 8·07; Cl,  $20\cdot422_{\infty}^{\circ}$ .

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# α-SPINASTEROL GLUCOSIDE AND OTHER CONSTITUENTS OF MAESA CHISIA\*

## ESAHAK ALI, V. S. GIRI and S. C. PAKRASHI†

Indian Institute of Experimental Medicine, Calcutta-700032, India

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**Key Word Index**—*Maesa chisia*; Myrsinaceae; fatty acids; fatty alcohols; long-chain ketones; α-spinasterol; α-spinasterol- $\beta$ -D-glucoside; stigmasta-8(14),22-dien-3 $\beta$ -ol;  $\beta$ -amyrin.

Plant. Maesa chisia D. Don. Source. Sub-Himalayan region. The specimen is available in the herbarium of Central Drug Research Institute. Lucknow. *Uses.* Insecticide [1]. *Previous work.* Nil.

Present work. Leaves and branchlets. Light petrol extracts on chromatography yielded (a) a mixture of long chain methyl ketones, mp 76-77° (IR 1720 cm<sup>-1</sup>, positive DNP test) with C-31 and

<sup>†</sup> Method B: calculated from amino acid analyser.

<sup>\*</sup> Part 33 in the series, "Studies on Indian Medicinal Plants". For Part 32 see Pakrashi, S. C., Achari, B. and Majumdar, P. C., *Indian J. Chem.* communicated.

<sup>†</sup> To whom correspondence should be addressed.